



HEALTH HOLDING

HAFER ALBATIN HEALTH  
CLUSTER

MATERNITY AND  
CHILDREN HOSPITAL

|                          |   |                         |                |
|--------------------------|---|-------------------------|----------------|
| <b>Department:</b>       | Laboratory and Blood Bank ( Parasitology) |                         |                |
| <b>Document:</b>         | Internal Policy and Procedure             |                         |                |
| <b>Title:</b>            | Microscopic Examination of Stool          |                         |                |
| <b>Applies To:</b>       | All Laboratory Staff                      |                         |                |
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## 1. PURPOSE:

- 1.1 To establish system and set responsibility for work.
- 1.2 To elucidate the proper procedure for stool examination.

## 2. DEFINITONS:

N/A

## 3. POLICY:

- 3.1 Stool sample volume should be at least 3gm.
- 3.2 Stool sample should be not contaminated with water or non-food debris, non-absorbable antidiarrheal drugs.
- 3.3 Microscopic examination of direct smear is done to assess the worm burden of patient
- 3.4 Microscopic examination of direct smear is done to provide quick diagnosis.
- 3.5 Microscopic examination of direct smear is done to check organism motility.

## 4. PROCEDURE:

### 4.1 Principle:

- 4.1.1 The microscopic examination of a direct smear has several purposes: to assess the worms burden of a patient, to provide a quick diagnosis of a heavily infected specimen, to check organism motility and to diagnose parasites that may be lost in concentrated techniques.

### 4.2 Specimen:

- 4.2.1 Any fresh stool specimen.

### 4.3 Quality control for direct smear:

- 4.3.1 Check the working iodine solution each time it is used.
- 4.3.2 Iodine should be the color of strong Orange Peko tea, discard if it is too light.
- 4.3.3 Protozoan stained with iodine should contain yellow gold cytoplasm, brown glycogen material and paler refractile nuclei. The chromatid bodies may not be as clearly visible as in a saline mount.
- 4.3.4 The microscope should be calibrated (within the last months).
- 4.3.5 Ensure that reagents and chemicals used are not expired.

### 4.4 Procedure:

- 4.4.1 Place one drop of 0.9% normal saline on the left side and one drop of iodine (working solution) on the wright side of the slide.
- 4.4.2 Take a small amount of fecal specimen and thoroughly emulsify the stool in saline and iodine using an applicator stick.
- 4.4.3 Slide a 22 mm cover slip at an angle into the edge of the emulsified faecal drop, push the cover slip across the drop before allowing it to fall into place .
- 4.4.4 Systematically check the whole 22 mm cover slip overlapping fields with 10 objective lenses.
- 4.4.5 Switch to high dry 40 objective lens for more details of any suspect eggs or protozoa.

#### 4.5 Interpretation:

- 4.5.1 Protozoan stained with iodine should contain yellow gold cytoplasm, brown glycogen materials and paler retractile nuclei.
- 4.5.2 The chromatoidal bodies may not be as clearly visible as in saline motion.
- 4.5.3 Protozoal trophozoite, cysts and helminth eggs and larva can be seen and identified.

#### 5. MATERIALS AND EQUIPMENT:

- 5.1 Normal saline (0.9%).
- 5.2 Lugols iodine solution 10%
- 5.3 Slides
- 5.4 Cover slips
- 5.5 Wooden sticks

#### 6. RESPONSIBILITIES:

- 6.1 The assigned technetium will perform the test







#### 7. APPENDICES:

N/A

#### 8. REFERENCES:

- 8.1 Cheesbrough, Monica District Laboratory Practice in Tropical Countries. Part 1: Pt.1 Press (1999)

#### 9. APPROVALS:

|              | Name                          | Title                              | Signature   | Date             |
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